

Epidemiology and Control of Alternaria Leaf Spot

Project No.: 07-PATH3-Adaskaveg

Project Leader: J. E. Adaskaveg
Dept. of Plant Pathology
University of California at Riverside
200 University Office Building
Riverside, CA 92521
(951) 827-7577
jim.adaskaveg@ucr.edu

Project Cooperators: H. Förster – University of California at Davis
G. Driever – University of California at Riverside
M. Viveros and P. Schrader - UCCE, Kern County

Objectives:

1. Etiology

- a. Identify pathogenic species of *Alternaria* using molecular methods.
- b. Determine the pathogen species composition within and between selected orchards at the beginning and at the end of the log phase of the epidemic.

2. Epidemiology

- a. Growth chamber studies to evaluate disease development under defined environmental conditions (wetness, relative humidity, temperature).
- b. Develop disease progress curves in relation to microclimates
 - i. Continue to evaluate environmental parameters (wetness, relative humidity, temperature) as they relate to disease development within and between orchards.
- c. Continue to evaluate the DSV model as a method of forecasting the disease with the goal of improved timing of fungicide treatments.
 - i. Evaluate 10- and 20-point thresholds of the 7-day index of the DSV model for predicting disease and timing of fungicides.
 - ii. Determine optimal fungicide application timings based on different methods for calculating thresholds of the DSV model.

3. Management

- a. Evaluate new fungicides for their efficacy in managing *Alternaria* leaf spot. Fungicides to be evaluated include selected strobilurin fungicides (Abound, Flint, Pristine) and non-strobilurin fungicides (e.g., Elite, difenoconazole – Inspire, polyoxin-D - Endorse, and other materials).
- b. Evaluate rotation programs that use calendar-based fungicide applications in comparison with applications based on the modified DSV model.
- c. Monitor for strobilurin field-resistance and shifts in sensitivity in populations of *Alternaria* spp. to other fungicides (e.g., boscalid).

Interpretive Summary:

Strobilurin- or Q_oI-resistant isolates of *Alternaria* spp. were first detected in 2003 and 2004, and crop losses (practical resistance) were common in 2006 in Kern Co. and again in 2007 in several almond-producing counties in California including (Kern, Glenn, Butte, and Tehama Co.). Once resistance occurs within a class (same mode of action), all fungicides within the class are also ineffective (i.e., azoxystrobin - Abound, trifloxystrobin - Gem, and pyraclostrobin - a component of Pristine) are ineffective once resistance develops. Still, Pristine was registered and recommended against *Alternaria* leaf spot with a 25-day PHI interval because the second fungicide, boscalid (carboxamide class), in the premixture was also active against the pathogen complex. The baseline sensitivity was determined for boscalid with EC₅₀ values of 0.015-0.058 ppm. In 2007, we detected carboxamide resistance for the first time in almond populations of *Alternaria* spp. in several counties including Kern, Glenn, and Butte Co. Resistant populations had EC₅₀ values of 10-50 ppm, an over 100-fold increase. Practical resistance is expected to be common next year if resistance spreads within the pathogen population similar as for the Q_oI fungicides. Thus, in order to replace the strobilurins and carboxamides with new fungicides for managing *Alternaria* leaf spot and other foliar diseases of almond, our studies focused on generating data to identify, characterize, and register new effective materials of different modes of action against the pathogens. The two new compounds that show the best promise are the DMI (demethylation inhibitor – Group 3) fungicide difenoconazole and the chitin synthase inhibitor (Group 19) polyoxin-D (a biofungicide). Baseline sensitivity data were developed for difenoconazole with an EC₅₀ value range of 0.01-0.05 ppm and is ongoing for polyoxin-D. Difenoconazole is being registered through the IR-4 program and polyoxin-D has been submitted into the program. In our 2007 trials in Kern and Colusa Co., three calendar-based applications of polyoxin-D (Endorse), difenoconazole (Inspire), or a SBI-strobilurin pre-mixture (Adament) were highly effective in reducing the incidence of disease from the control and they performed similar to or significantly better than multiple applications of Pristine, Captan, or Scala. Rovral was also highly effective in these trials; however, the EPA has indicated that they will not change the current registration of this fungicide on almond. Growth chamber experiments under controlled temperature-wetness conditions were repeated and demonstrated that only plants with mite injuries developed disease symptoms in these studies. Thus, injuries from mites and potentially other insects are an important focus in an integrated management strategy for *Alternaria* leaf spot.

Introduction. *Alternaria* leaf spot of almond is a disease that is caused by three species in the *Alternaria alternata* complex, *A. arborescens*, *A. alternata*, and *A. tenuissima*. Under favorable conditions for disease development, trees can be completely defoliated by early to mid-summer. The disease occurs throughout the almond production areas in the central valleys of California. It is severe almond production areas where dew forms, the air is stagnant, and temperatures are high or in areas where almonds have been planted in poorer soils and irrigation practices require more frequent and extended watering into the summer. In the last several years, serious disease outbreaks in northern and southern growing areas have been reported.

Because the occurrence of *Alternaria* leaf spot of almond is greatly influenced by microclimatic conditions such as temperature and wetness within orchards, the Disease Severity Value (DSV) model that is used for forecasting of *Alternaria* diseases on other crops was evaluated on almond in our previous years' studies. We demonstrated a close correlation between the increase in actual disease and increases in DSV that are determined by the number of hours of wetness within defined temperature ranges. In subsequent studies, we adjusted threshold temperatures to ≥ 62 F to refine the accuracy of the model for almond in California. In addition, we demonstrated that the actual development of disease incidence correlated with environmental conditions that occurred 25 days (± 3 days) before the onset of disease symptoms (the disease progress curve latency period). Thus, temperature-leaf wetness conditions used for calculation of the DSVs reflect the actual disease development. The DSV model is a tool for predicting infection periods for the *Alternaria* pathogen on almond and for timing of fungicide applications (This model was used for fungicide applications in 2007).

Progress on new and amended fungicide registrations. In 2007, Captan and Scala were registered under 2ee amendments for *Alternaria* leaf spot, but both only have suppressive action based on our evaluations. The label for Rovral, however, still only includes up to 5-wk after petal fall applications and EPA is unwilling to change the PHI based on our Section 18 submission in 2007. Pristine was labeled with a 25-day PHI in 2006. With the widespread occurrence of strobilurin (Q_oI) resistance and recent identification of boscalid resistance (see below) in populations of *Alternaria* spp. on almond, however, practical (field) resistance and crop loss has subsequently occurred.

First evaluated by us, the two new fungicides Inspire (difenoconazole) and Endorse (polyoxin-D) continued to be the most effective treatments in our 2007 trials. Difenoconazole is a sterol biosynthesis inhibiting (SBI) fungicide that is registered in other countries for the control of diseases caused by *Alternaria* spp., whereas polyoxin is a chitin-inhibiting biofungicide that is registered in the US on turf and in other countries for the control *Alternaria* diseases on several crops. Difenoconazole is being registered through the IR-4 program and the field and lab portions were completed in 2006 and 2007, respectively. Submission to EPA is planned in 2007/08 and registration on almond is expected in 2009/10. Arysta Life Science has also requested a registration of polyoxin-D at 50-100 g ai/A on almond pending EPA review. Both fungicides were also recommended for emergency registration in 2008.

Growth chamber studies to evaluate disease development under defined environmental conditions. After 10-14 days of incubation, *Alternaria* leaf spot symptoms and sporulation of the fungus only developed on mite-injured plants, but not on healthy plants, and all three species of the pathogen caused disease in these growth chamber studies. Thus, for a second year, we demonstrated that mite-injured almond leaves are much more susceptible to *Alternaria* leaf spot. Field observations in 2007 also indicated that with less mite injury, the disease was less severe. Thus, mite or other types of leaf injury may increase the severity of *Alternaria* leaf spot outbreaks on almond.

Development of baseline sensitivity data and resistance assessment studies. The baseline sensitivity of *Alternaria* spp. from almond that were never exposed to

the fungicide boscalid consisted of a range of EC₅₀ values from 0.015-0.058 ppm. In 2007, we detected carboxamide resistance for the first time in almond populations of *Alternaria* spp. in several counties including Kern, Glenn, and Butte Co. Resistant populations had EC₅₀ values of 10-50 ppm, an over 100-fold increase. Practical or field resistance is expected to be common in the next year if resistance spreads within the pathogen population similar to Q_oI fungicides. Thus, in order to replace the strobilurins and carboxamides with new highly effective fungicides for managing *Alternaria* and other foliar diseases of almond, we identified two new effective materials of different modes of action: the

Table 1. Efficacy of fungicide treatments for management of *Alternaria* leaf spot on almond - Kern Co. 2007

A. Trial 1 - Carmel

| Product | Rate | Incid. (leaves) | | Defoliation | |
|-------------------------------|-----------------|-----------------|-----|-------------|------|
| | | (%) | LSD | Rating** | LSD |
| Control | -- | 78.9 | a | 2.7 | a |
| Inspire 250EC | 5 fl oz | 18.0 | bcd | 1.1 | bcd |
| Inspire 250EC | 7 fl oz | 16.6 | bcd | 0.5 | e |
| Orbit 3.6EC | 6 fl oz | 27.4 | bc | 0.9 | bcde |
| Polyoxin D 11.2WG | 50 g ai | 14.1 | cd | 0.5 | e |
| Polyoxin D 11.2WG | 100 g ai | 11.9 | d | 0.6 | de |
| Rovral 4F | 16 fl oz | 21.8 | bcd | 1.3 | bc |
| Rovral 4F | 32 fl oz | 12.8 | cd | 0.9 | bcde |
| Inspire + Captan 80WDG | 7 fl oz - 5 lb | 23.7 | bcd | 1.4 | b |
| Rovral 4F + Captan 80WDG | 32 fl oz - 5 lb | 19.5 | bcd | 0.7 | bd |
| Pristine 38 WG + Captan 80WDG | 14.5 oz - 5 lb | 31.4 | b | 1.3 | bc |
| Inspire Super | 14 fl oz | 28.1 | b | 0.8 | cde |
| A13703(I-V) | 10 fl oz | 22.4 | bcd | 1.0 | bcde |
| Pristine 38WG | 14.5 oz lb | 18.2 | bcd | 0.9 | bcde |

B. Trial 1 - Monterey

| Product | Rate | Incid. (leaves) | | Defoliation | |
|----------------------------|----------------|-----------------|-----|-------------|------|
| | | (%) | LSD | Rating** | LSD |
| Control | --- | 80.0 | a | 3.2 | a |
| Inspire 250EC | 7 fl oz | 21.4 | de | 0.7 | fg |
| Orbit 3.6EC | 6 fl oz | 17.6 | e | 1.7 | bcd |
| Polyoxin D 11.2WG | 50 g ai | 16.6 | e | 1.6 | bcd |
| Polyoxin D 11.2WG | 100 g ai | 15.7 | e | 0.9 | efg |
| Rovral 4F | 32 fl oz | 23.6 | de | 1.1 | def |
| Inspire + Captan 80WDG | 7 fl oz - 5 lb | 49.2 | b | 1.9 | bc |
| Orbit 3.6EC + Captan 80WDG | 6 fl oz - 5 lb | 36.7 | bc | 2.1 | b |
| Inspire Super | 11.5 fl oz | 26.3 | cde | 1.3 | cdef |
| Inspire Super | 14 fl oz | 16.8 | e | 1.7 | bcd |
| Inspire Super | 20 fl oz | 25.6 | cde | 1.4 | cde |
| A13703(I-V) | 10 fl oz | 27.6 | cde | 0.5 | g |
| Adament 50WG | 8 oz | 31.8 | cd | 1.4 | cde |
| Pristine 38WG | 14.5 oz lb | 21.2 | de | 1.8 | bc |

C. Trial 2 - Monterey

| Product | Rate | Incid. (leaves) | | Defoliation | |
|----------------------------------|-----------------|-----------------|-----|-------------|-----|
| | | (%) | LSD | Rating** | LSD |
| Control | --- | 75.2 | a | 3.3 | a |
| Polyoxin D 11.2WG + Captan 80WDG | 6.3 oz - 5 lb | 44.8 | b | 2.7 | b |
| Scala 600SC + Captan 80WDG | 18 fl oz - 5 lb | 50.4 | b | 2.5 | b |
| Vanguard 75WG + Captan 80WDG | 10 oz - 5 lb | 41.2 | b | 2.6 | b |

* - Trees were treated with or without Captan at 5-wk-after petal fall (grower applied). Treatments were applied using an air-blast sprayer at 100 gal/A and there were 4 single-tree replications. In Tables A & B, applications were on 5/8, 5/31, and 6/25/07. In Table C, applications were on 5/31 and 6/25/07.

** - Evaluations were done on 8-9-07. For disease incidence on leaves, 200 leaves from each of the 4 single-tree reps were evaluated for the presence of disease. For evaluation of tree defoliation on 9-12-07, trees were rated based on a scale from 0 (= full canopy), 1 (<25% defoliation) to 4 (100% defoliation). Values followed by the same letter are not significantly different based on an ANOVA and LSD mean separation ($P > 0.05$).

DMI(demethylation inhibitor – Group 3) fungicide difenoconazole and the chitin synthase inhibitor (Group 19) polyoxin-D (a biofungicide). All isolates evaluated were sensitive to difenoconazole and EC₅₀ values for inhibition of mycelial growth ranged from 0.01 to 0.05 ppm. Research is ongoing for polyoxin-D.

Fungicide evaluations for management of *Alternaria* leaf spot of almond in 2007.

In the first trial, the five-week-after-petal-fall application of Captan did not significantly improve disease management as compared to no petal fall fungicide treatment and there was no interaction with the late spring applications of fungicides. Thus, data from the two main plots were combined. The late spring applications, however, were significant treatments in reducing *Alternaria* leaf spot. All of the treatments significantly reduced disease from 78.9% and 80% in the untreated controls to less than 30% and less than 50% incidence on cvs. Carmel and Monterey, respectively (Table 1A,B). Defoliation ratings were reduced from 2.7 and 3.2 to less than 1.3 and 2.1 on cvs. Carmel and Monterey, respectively. The best treatments in reducing the incidence of disease and defoliation were polyoxin-D, Rovral, and Inspire on both cultivars (Table 1A,B). Pristine also significantly reduced the disease on both cultivars although a high level of strobilurin resistance was detected in the *Alternaria* spp. population in this orchard with over 80% of the isolates being resistant. The efficacy of Pristine can be explained by the relatively low incidence of boscalid resistance in pathogen populations of this orchard (ca. 30%) and therefore, rendering this fungicide still effective in reducing the disease. With the further spread of boscalid resistance due to over-use of Pristine, however, efficacy of this fungicide will most likely continue to decline. Our data also indicate that the addition of captan to selected fungicides was not beneficial for disease control and sometimes decreased the performance when used in tank mixtures. The addition of other fungicides also did not improve the performance of Inspire (Table 1A,B). Late spring, early summer applications of Polyoxin-D, Scala, or Vanguard that were all mixed with Captan significantly reduced the disease, but these treatments were not as effective as the earlier timings (Table 1C).

Thus, we identified the most effective treatment timing for management of *Alternaria* leaf spot to be late spring applications that are best done following the temperature-wetness-based DSV model. The petal fall application with Captan did not reduce the disease; however, when we used a mixture of Captan with Rovral at this timing in last year's trials, there was a significant reduction in disease. This implies that Rovral is a more effective fungicide against *Alternaria* leaf spot than Captan. Rovral was not used as a petal fall treatment in 2007 because we had submitted this fungicide to be re-labeled to allow late spring treatments, and we successfully used Rovral this year at these timings. With EPA not approving this re-labeling, Rovral should best be used in petal fall treatments that are followed by late-spring applications with other fungicides to obtain a high level of disease control. Although Captan is not very effective against *Alternaria* spot, a mixture of Rovral with Captan will have the benefit to be effective against scab as well.

In a field trial in Colusa Co., Bravo was applied at petal fall to reduce scab. With a lower level of *Alternaria* leaf spot in the untreated control as compared to the Kern Co. trial, all

treatments when applied between late April and early June were very effective in managing the disease (Table 2A). Pristine was also effective at this location where boscalid resistance was not found. Late spring applications with Scala, Captan, or mixtures of Captan with Scala or Polyoxin were less effective and subsequently resulted in high levels of tree defoliation (Table 2B).

Table 2. Efficacy of fungicide treatments for management of *Alternaria* leaf spot on almond cv. Carmel - Colusa Co. 2007

A. Trial 3 - Carmel

| Product | Rate | Incid. (leaves) | | Defoliation | |
|--------------------------|-----------------|-----------------|-----|-------------|-----|
| | | (%) | LSD | Rating** | LSD |
| Control | --- | 31.6 | a | 2.50 | ab |
| Inspire 250EC | 7 fl oz | 3.9 | bc | 0.88 | d |
| Orbit 3.6EC | 6 fl oz | 3.5 | c | 1.63 | bcd |
| V-10116 | 4 fl oz | 5.0 | bc | 1.13 | dc |
| Polyoxin D 11.2WG | 50 g ai | 9.4 | b | 1.13 | dc |
| Polyoxin D 11.2WG | 100 g ai | 2.6 | c | 1.88 | bcd |
| Rovral 4F | 32 fl oz | 6.1 | bc | 1.50 | bcd |
| Inspire + Vanguard 75WG | 7 fl oz - 5 lb | 5.6 | bc | 1.25 | bd |
| Rovral 4F + Captan 80WDG | 32 fl oz - 5 lb | 3.0 | bc | 2.88 | a |
| Pristine + Captan 80WDG | 6 fl oz - 5 lb | 4.8 | bc | 2.63 | ab |
| Adament 50WG | 8 oz | 5.3 | bc | 2.50 | ab |
| Pristine 38WG | 14.5 oz lb | 2.6 | c | 1.38 | bcd |

B. Trial 4 - Carmel

| Product | Rate | Incid. (leaves) | | Defoliation | |
|----------------------------|-----------------|-----------------|-----|-------------|-----|
| | | (%) | LSD | Rating** | LSD |
| Control | --- | 46.6 | a | 3.63 | a |
| Scala 600SC | 18 fl oz | 16.9 | b | 2.88 | ab |
| Captan 80WDG | 5 lb | 16.3 | b | 2.63 | b |
| Scala 600SC + Captan 80WDG | 18 fl oz - 5 lb | 21.6 | b | 2.88 | ab |
| Polyoxin D + Captan 80WDG | 10 oz - 5 lb | 21.0 | b | 3.13 | ab |

* - Trees were treated with Bravo at 5-wk-after petal fall (applied by the grower). Treatments were applied using an air-blast sprayer at a rate of 100 gal/A. There were 4 single-tree reps for each sub-plot treatment. In Table A, treatments were applied on 4/25, 5/18, & 6/6/07. In Table B, treatments were applied on 6/6 and 6/21/07.

** - Evaluations were done on 8-15-07. For disease incidence on leaves, 200 leaves from each of the 4 single-tree reps were evaluated for the presence of disease. For evaluation of tree defoliation on 9-12-07, trees were rated based on a scale from 0 (= full canopy), 1 (<25% defoliation) to 4 (100% defoliation). Values followed by the same letter are not significantly different based on an ANOVA and LSD mean separation ($P > 0.05$).

In summary, management of *Alternaria* leaf spot with fungicides as part of an integrated strategy should start with petal fall applications that include Rovral followed by late spring applications with other materials. Because no effective materials are currently registered, registration of Inspire (difenoconazole) and the biofungicide polyoxin-D will be instrumental in the management of *Alternaria* leaf spot of almond. These materials will have to be strictly used in rotations or mixtures to delay the development of resistance and maintain their efficacy. Also, the pre-mixture Adament performed well, but concerns exist because one of the components of this product is a strobilurin fungicide. Additional materials with activity against *Alternaria* spp. need to be identified.

Because of the current limited arsenal of chemical treatments available, other components of an integrated approach in disease management are even more critical.

These include insect and mite control as well as cultural practices that lead to a decreased humidity in the micro-environment in the orchard including hedging, improvement of water penetration into the soil by adding gypsum, and changing the watering or irrigation schedule to less frequent irrigation using soil moisture probes. If all components of the disease triangle are considered, fungicide treatments will be the most beneficial.