

Orchard Weed Management Research

Brad Hanson^{1,2}, Seth Watkins¹, Marcelo Moretti¹, Lynn Sosnoskie¹, Bahar Yildiz Kutman¹, Sarah Morran¹, Caio Brunharo¹, Mariano Galla¹, Lucas Bobadilla¹, Franz Niederholzer², John Roncoroni², and David Dol²
¹Department of Plant Sciences, University of California, Davis; ²University of California Cooperative Extension

Objectives

The overall goals of the tree and vine weed science research and extension program at UC Davis (<http://ucanr.org/brad.hanson>) is to provide information on weed management and herbicide issues to California growers, Pest Control Advisors, and the UC Cooperative Extension network. The almond industry is one of the key stakeholder groups for this program; however, the majority of our research is broadly applicable to, and partially supported by, other orchard and vineyard commodities in the state as well as the forest control industry.

The specific objectives of this ongoing project (14Hort12.Hanson – Weed Management) mirror the major research areas in our program:

- Evaluation and testing of newly registered materials, tank mix partners, and application techniques for control of weeds with a special focus on glyphosate-resistant species.
- Evaluating herbicide injury symptoms in almonds and developing training tools for Farm Advisors and pest control industry advisors and consultants.

Numerous field and greenhouse experiments were conducted in 2013/2014 to support grower, Pest Control Advisor, and Farm Advisor weed and herbicide research needs. Because a more thorough presentation of these projects is available in the annual reports to the Almond Board and in various online venues, this poster presents only a few representative results. Data from related projects are routinely presented at cooperative extension meetings as well as scientific presentations by members of the research team.



Herbicide performance

Several field trials were conducted to evaluate weed control efficacy in commercial almond orchards. In these experiments, research personnel applied replicated, small-plot treatments using CO2 pressurized backpack or ATV-mounted spray equipment. Weed control was visually assessed several times during the growing season and, in some cases, biomass or other quantitative data were collected. A few representative data are shown in Tables 1-5 below; a full accounting is available in the Almond Board Research Report. Many of these data are also presented online at the UC Weed Science blog (<http://ucanr.edu/blogs/UCDWeedScience/index.cfm>) and the Almond Doctor blog (<http://thealmonddoctor.com/>)

Table 1. Selected weed control evaluations from 2014-15 large plot demonstration conducted in an almond orchard near Escalon, CA; second year treatments. (Watkins and Hanson)

Table 2. Postemergence weed control in an almond orchard trial conducted near Wasco, CA in spring 2014. (Moretti, Watkins, and Hanson)

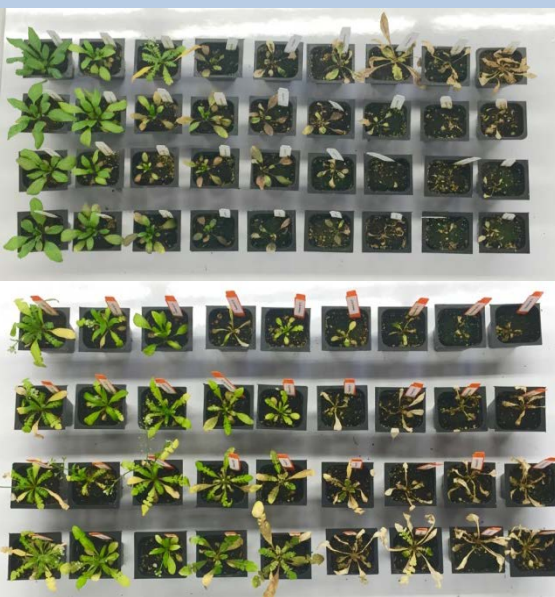
Table 3. Selected weed control evaluations from 2014-15 Broadworks and Alon tankmix trial near Arbuckle, CA (Brunharo and Hanson)

Table 4. Selected weed control evaluations from 2014-15 comparison of Alon and other preemergence tankmix and sequential partners in an almond orchard near Escalon, CA. All treatments included a high rate of Relay 280 and Roundup Powermax to ensure good control of existing weeds. (Watkins and Hanson)



Identification and verification of herbicide-resistant weeds

Weed management in California tree and vine crops is currently dominated by problems with glyphosate-resistant and glyphosate-tolerant species. To date, six species resistant to glyphosate have been confirmed: hairy fleabane, horseweed (aka mareetail), Italian and rigid ryegrass, and junglerice, and annual bluegrass. Several other species of concern have been identified and are under evaluation; these include three-spike goosegrass, Palmer amaranth and a suite of summer annual grasses. Research being conducted on herbicide-resistant weeds includes confirmation of resistance (photo at left), determining distribution of the resistant populations, evaluation of alternative control measures, and determining the underlying physiological and genetic causes of resistance.



Dose-response screening of suspected glyphosate-resistant shepherd's purse

Fleabane biology and control

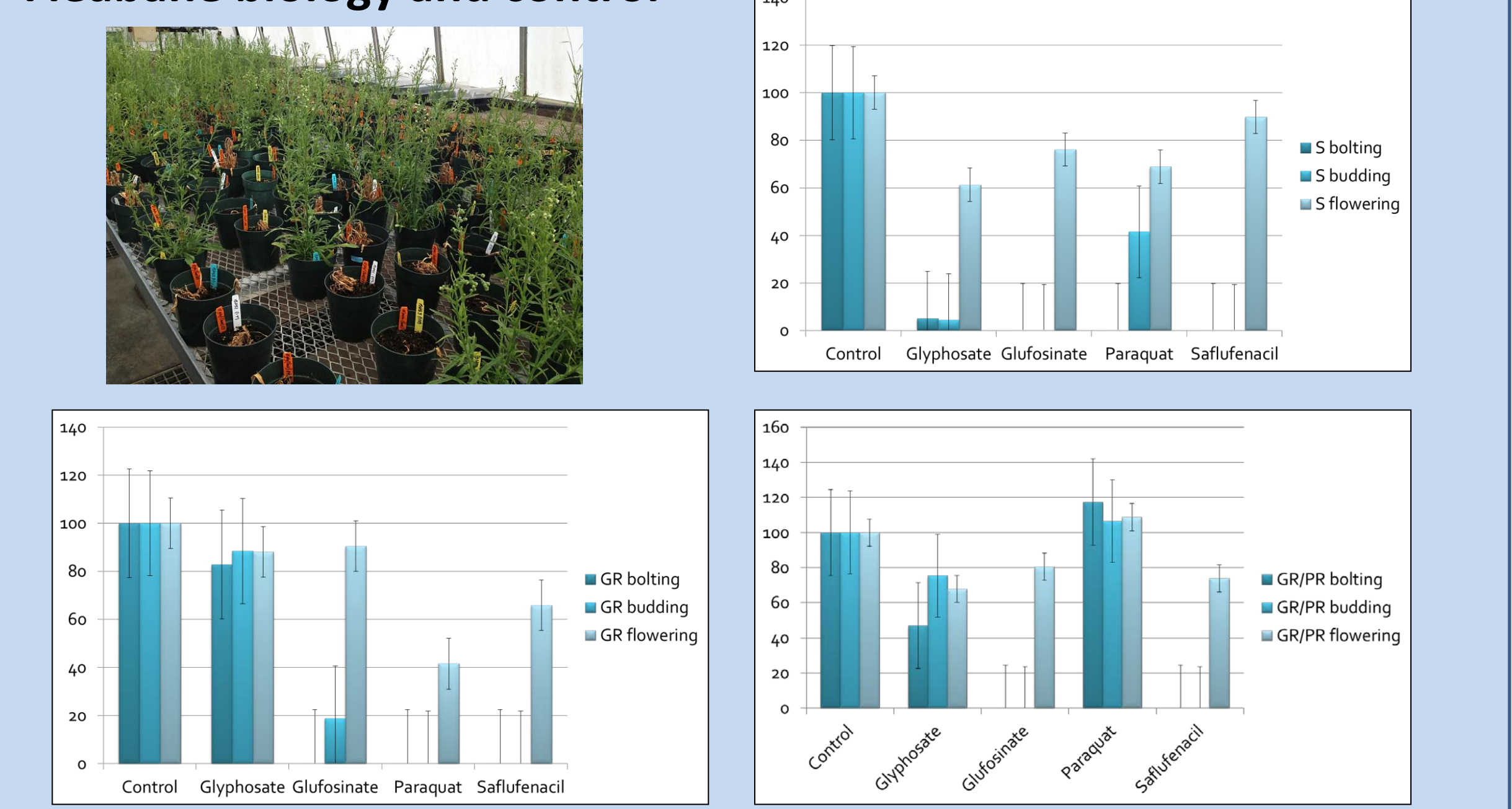


Figure 1. Biomass production (regrowth) of glyphosate susceptible (S), glyphosate-resistant (GR) and glyphosate-paraquat-resistant (GR/PR) hairy fleabane to four POST herbicides applied at early bolting, budding, or flowering growth stages. Once hairy fleabane reaches the flowering stage, none of the herbicides provided full control of any biotype (Sosnoskie and Hanson).

Junglerice biology and genetics

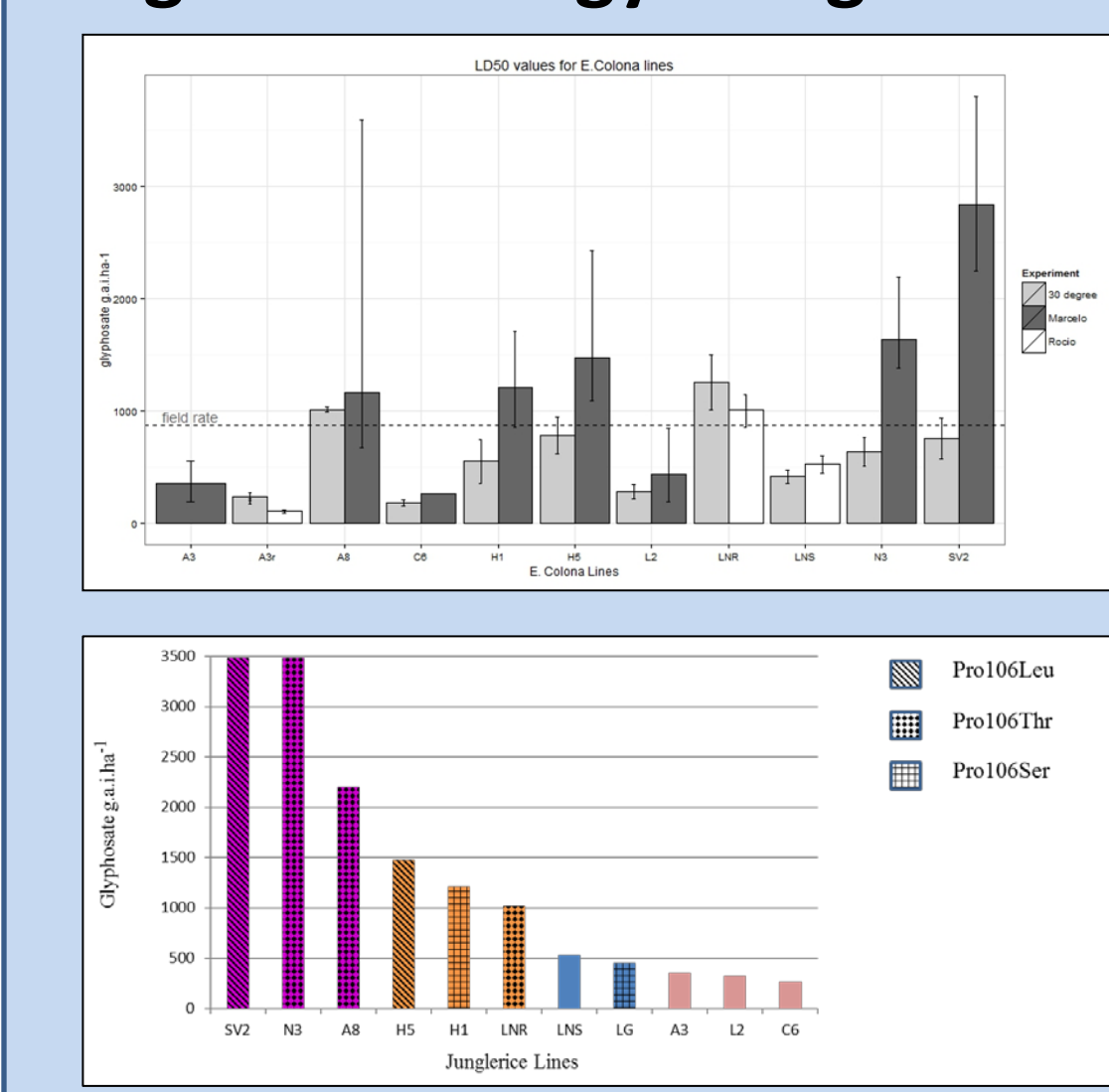


Figure 2. Glyphosate-resistant junglerice plants from the Central Valley have different EPSPS mutations that are associated with varying levels of glyphosate resistance at the whole plant level. Data indicate GR50 levels in greenhouse experiments on populations derived from previous field collections (Morran, Moretti, Fischer, and Hanson).

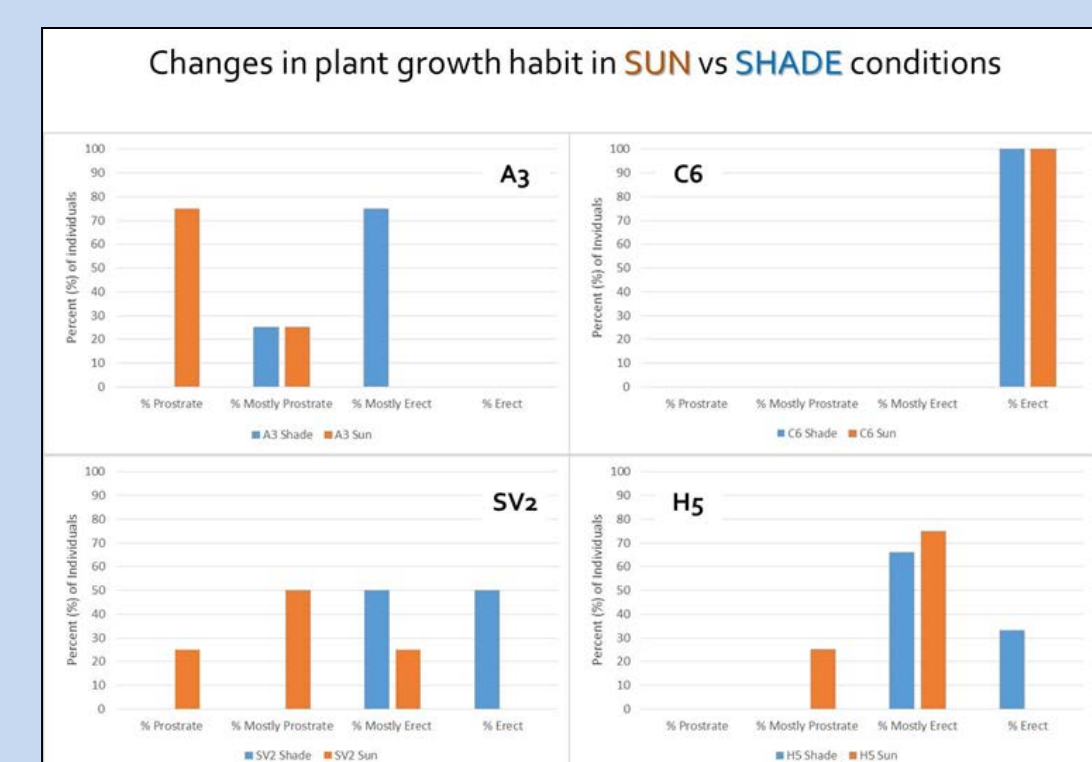


Figure 3. Glyphosate-resistant junglerice populations from various regions of California have different growth habits and responses to environmental cues (e.g. shade). Current research is ongoing to determine if or how this may be related to resistance to herbicides or other stresses and if the resistant populations are also more invasive than the wild types. (Sosnoskie, Morran, and Hanson).

Recent Publications

- Okada, M, B.D. Hanson, K.J. Hembree, Y. Peng, A. Shrestha, C.N. Stewart, S.D. Wright, and M. Jasienuk. 2014. Glyphosate resistance is more widespread in *Conyza bonariensis* than in closely related *C. canadensis* in California. Weed Research 55:173-184
- Hanson, B., S. Wright, L. Sosnoskie, A. Fisher, M. Jasienuk, J. Roncoroni, K. Hembree, S. Orloff, A. Shrestha, and K. Al-Khatib. 2014. Herbicide resistant weeds challenge signature California cropping systems. California Agriculture 68:142-152
- Hanson, B.D., A.J. Fischer, A. McHughen, M. Jasienuk, A. Shrestha, and A.J. Jhala. 2014. Herbicide-resistant weeds and crops. In S.A. Fennimore and C. Bell (eds). Principles of Weed Control. California Weed Science Society, Salinas CA (ISBN: 978-0-692-30482-2).
- Hanson, B.D., J. Roncoroni, K.J. Hembree, R. Molinar, and C.L. Elmore. 2014. Tree, vine, and soft-fruit crops. In S.A. Fennimore and C. Bell (eds). Principles of Weed Control. California Weed Science Society, Salinas CA (ISBN: 978-0-692-30482-2).
- Hanson, B.D. and T.W. Miller. 2015. The importance of 2,4-D in orchard, vineyard, and berry crops in the United States. Chapter in Phenoxy Herbicide task force report book (in press).
- Nguyen, T.T., D.C. Slaughter, B.D. Hanson, A. Barber, A. Freitas, D. Robles, and E. Whelan. 2015. Automated mobile system for accurate outdoor tree crop enumeration using an uncalibrated camera. Sensors 15:18427-18422.
- Moretti, M., A. Shrestha, K.J. Hembree, and B.D. Hanson. 2015. Postemergence control of glyphosate-paraquat resistant hairy fleabane (*Conyza bonariensis*) in tree nut orchards in the Central Valley of California. Weed Technol. 29:501-508.
- Moretti, M.L., L.M. Sosnoskie, A. Shrestha, S.D. Wright, K.J. Hembree, M. Jasienuk, and B.D. Hanson. 2015. Distribution of *Conyza* sp. In orchards of California and response to glyphosate and paraquat. Weed Technol. (in press)

Other support

In addition to support from the Almond Board of California, the UC Davis Weed Science Program is supported by other commodity boards, federal and state grant programs, and funding from the agricultural chemical industry. A special thanks to all of our sponsors, collaborators, and cooperators.

Multiple-resistant Conyza spp.

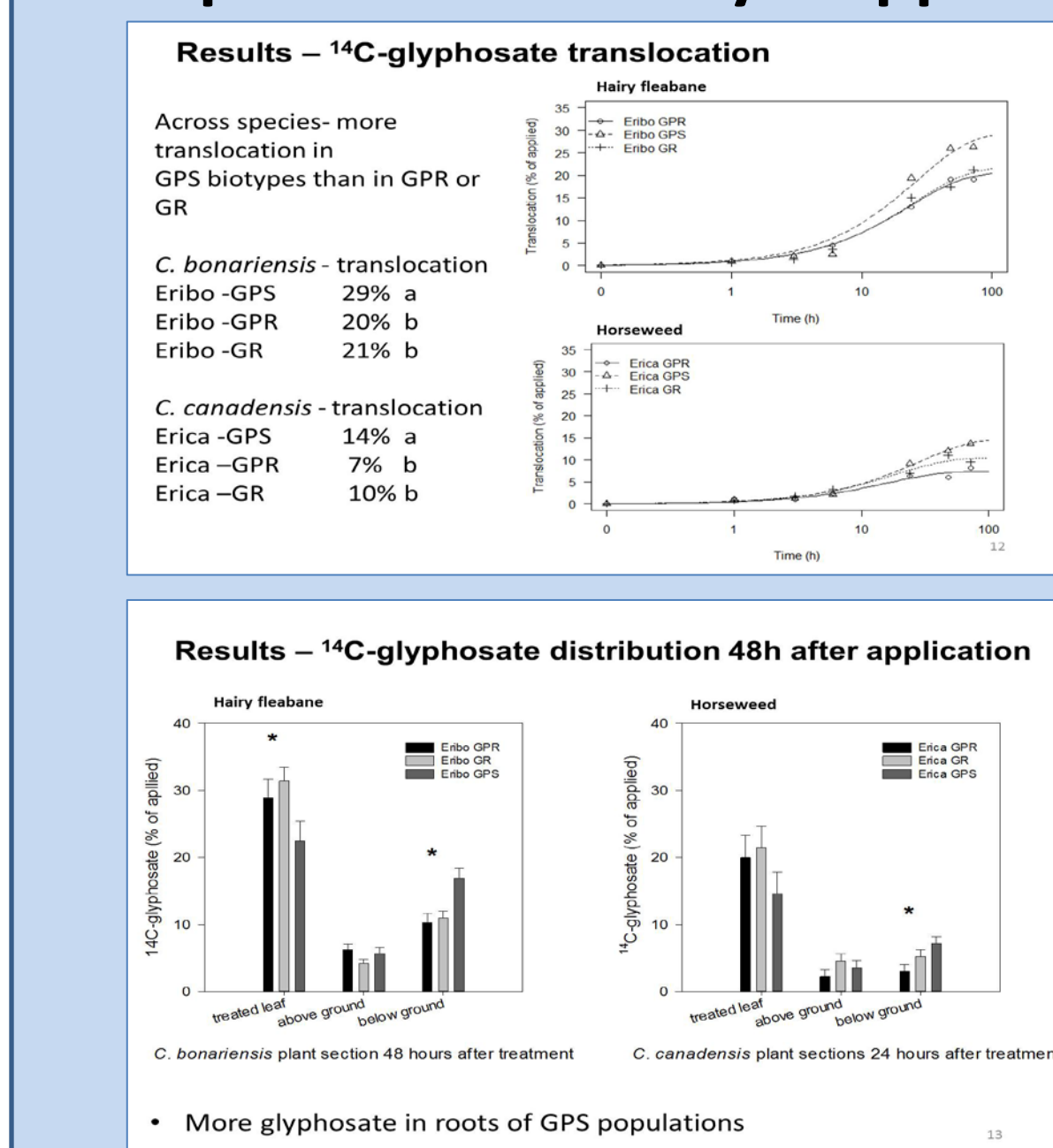


Figure 4. Evaluation of glyphosate translocation in glyphosate-susceptible (GPS), glyphosate-resistant (GR) and glyphosate-paraquat-resistant (GPR) hairy fleabane using radio-labeled glyphosate (Moretti and Hanson).

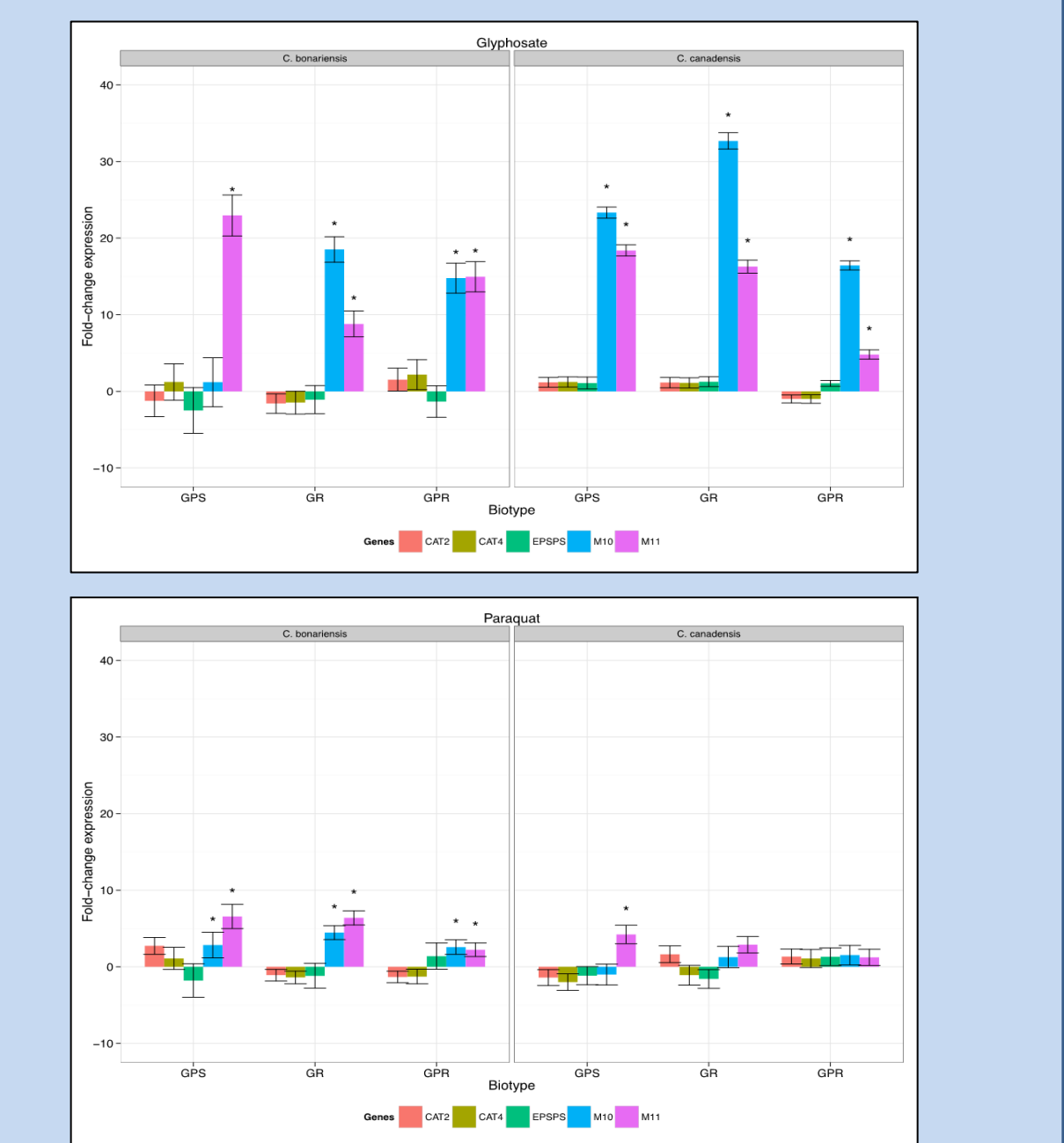


Figure 5. Evaluation of transcription of ABC transporter genes in response to glyphosate or paraquat in GPS, GR, or GPR hairy fleabane (*C. bonariensis*) and horseweed (*C. canadensis*) (Moretti, Morran, and Hanson).

Herbicide injury and crop safety

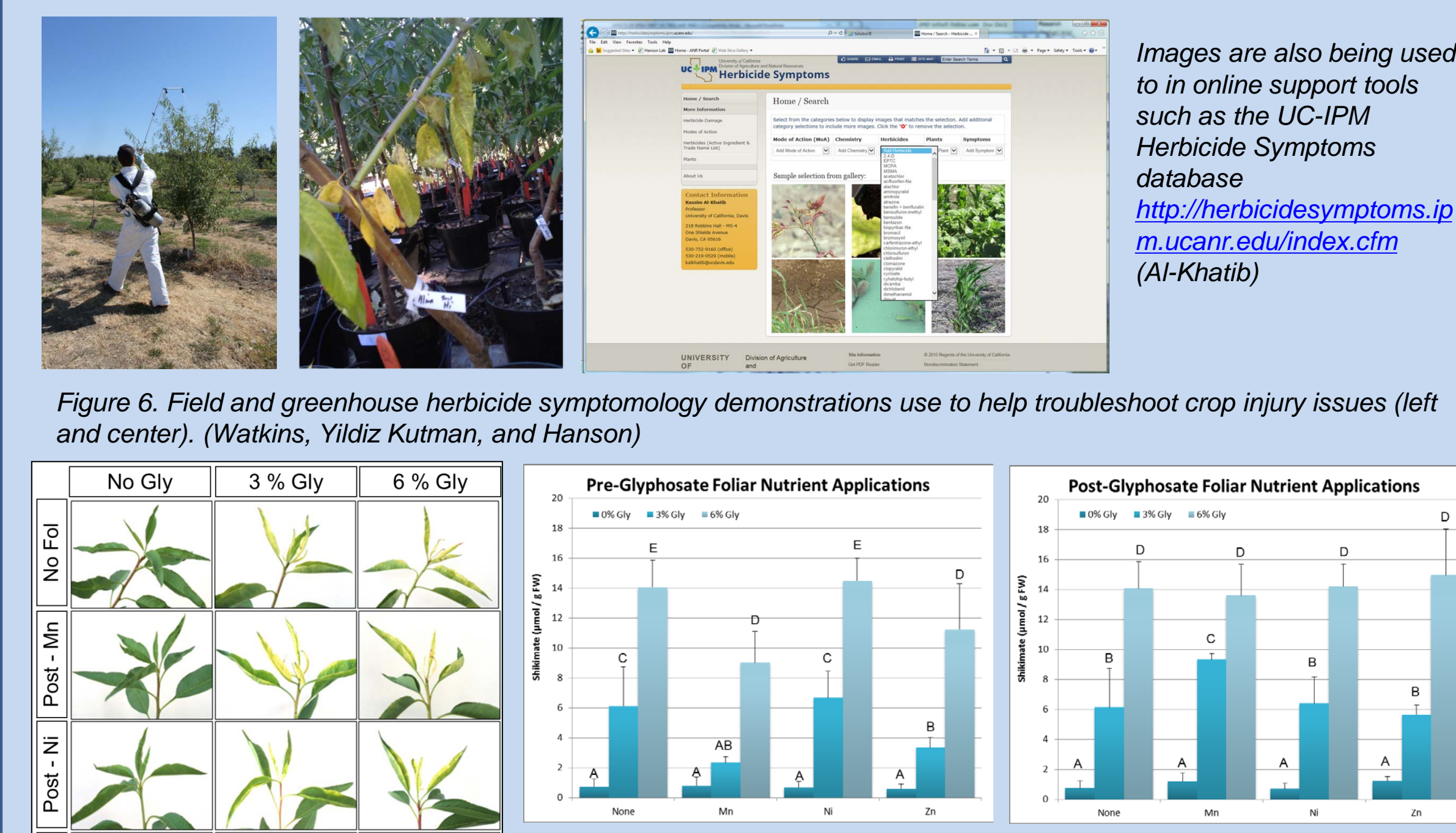


Figure 6. Field and greenhouse herbicide symptomatology demonstrations use to help troubleshoot crop injury issues (left and center). (Watkins, Yildiz Kutman, and Hanson)

Images are also being used to in online support tools such as the UC-IPM Herbicide Symptoms database <http://herbicidesymptoms.ipm.ucanr.edu/index.cfm> (Al-Khatib)

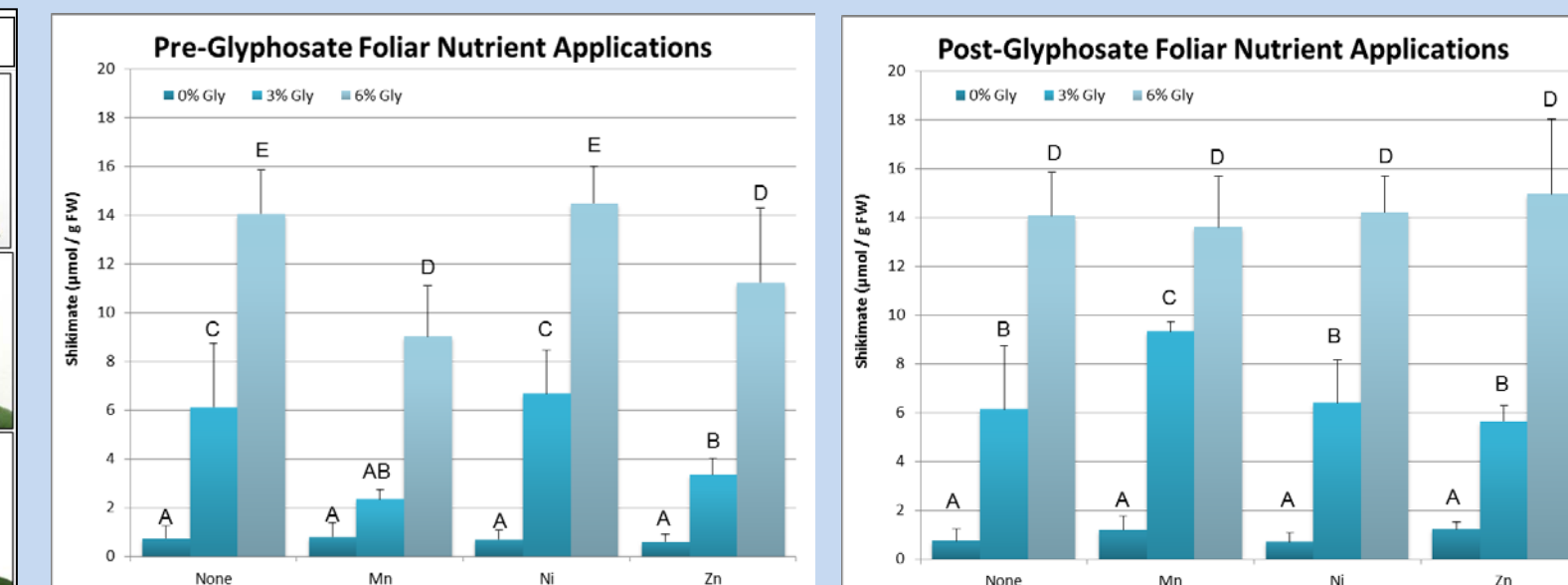


Figure 7. Greenhouse evaluation of micronutrient and glyphosate interactions. Glyphosate was applied at 0.03X and 0.06X of a normal use rate and trees were treated with micronutrient solutions either before or after glyphosate exposure. (Yildiz Kutman and Hanson).