

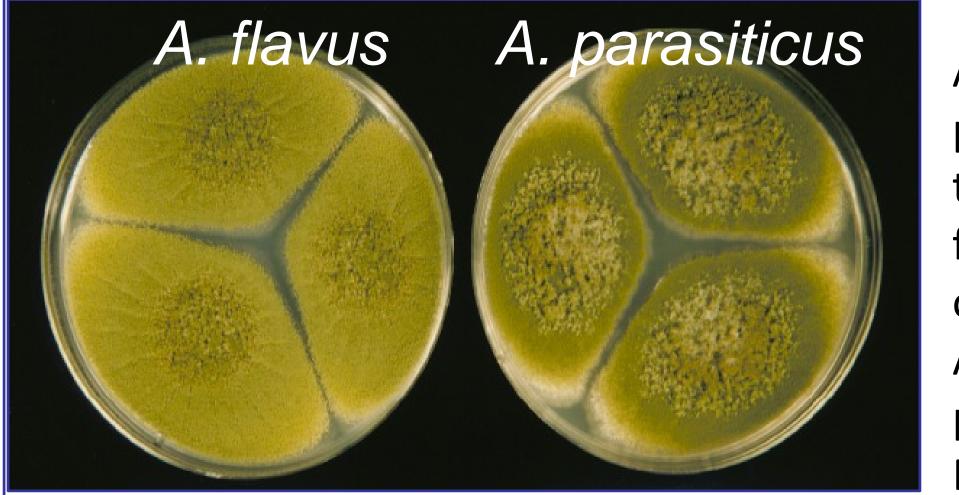
## Inoculum monitoring of biological control agent AF36, Aspergillus flavus, under field conditions



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## INTRODUCTION

Aflatoxins, produced by Aspergillus flavus and A. parasiticus (Fig. 1), are among the most potent natural carcinogens and are widely regulated by governments who have set very low tolerances for aflatoxins in food and animal feeds. California nut crops and other crops such as fig can be contaminated with aflatoxins (Fig. 2). During the last decade, we have been working on reducing aflatoxin contamination of nuts and figs using the atoxigenic strain of A. flavus AF36. Fortunately, the biological control A. flavus AF36 Prevail<sup>®</sup> has been approved for use in pistachio (2012), almond (2017), and fig (2017) orchards in California by the Environmental Protection Agency (EPA) and the California Department of Pesticide Regulation. Here, we present different field studies aimed to understand the dynamic of the atoxigenic A. flavus AF36 inoculum under field conditions in California.

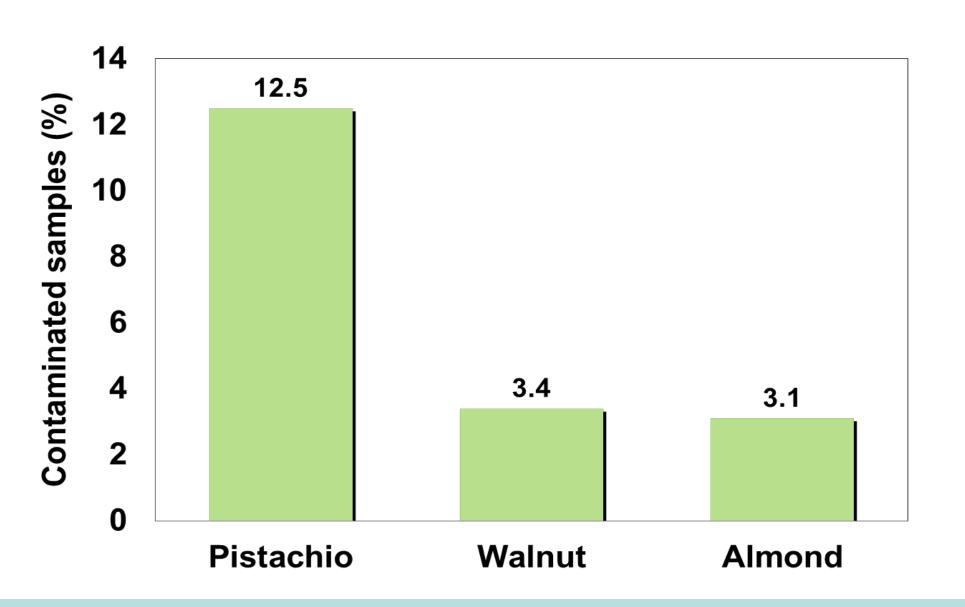
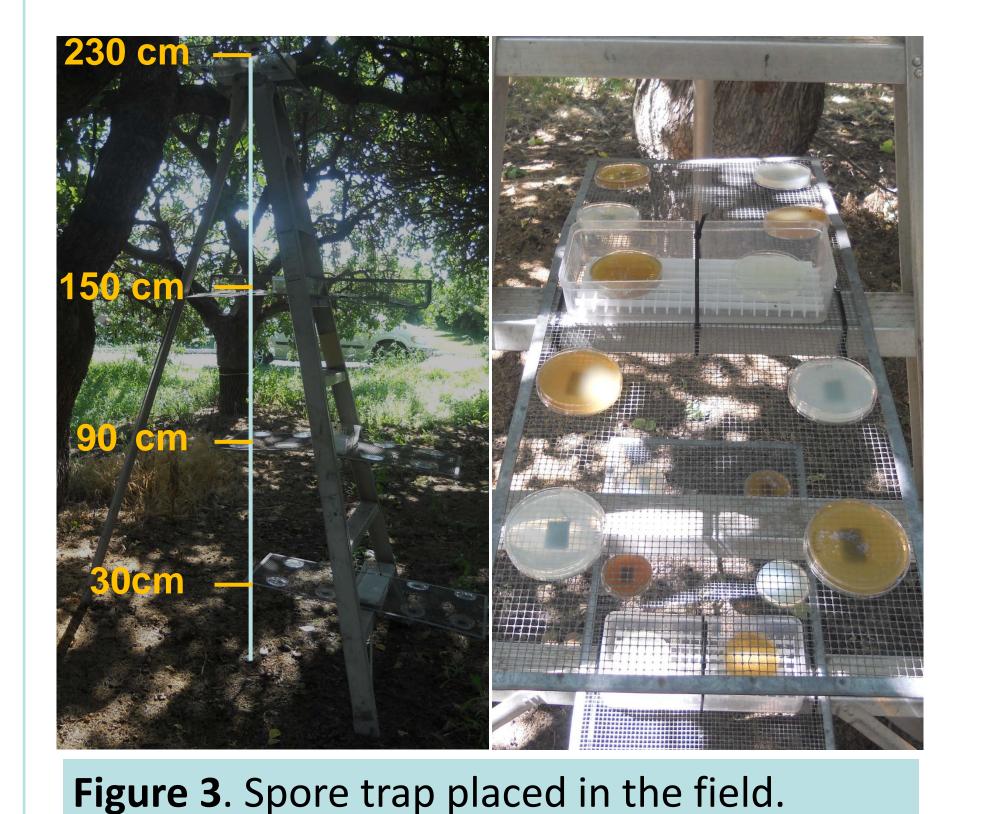


Figure 1. Fungi that can produce aflatoxins in almond orchards in California.

Figure 2. Californian nut samples contaminated with aflatoxins (data by the Dried Fruit Association).



## **PROCEDURES AND RESULTS**

*Effect of height on A. flavus AF36 spore density.*, The effect of height above the ground on spores density of A. flavus AF36 was studied under field conditions in an orchard at the Kearney Agricultural Research and Extension Center. Spore traps, consisting of eight 9-cm Petri dishes with 50 ml Aspergillus Differentiation Agar medium (Fluka, AFPA) per plate, were placed at different heights from 30 to 230 cm. The biocontrol product A. flavus AF36 Prevail<sup>®</sup> was applied on the soil under the spore traps (Fig. 3). In general, results indicate that A. flavus spores decreased exponentially as a function of height, while the density of A. niger spores increased (Fig. 4). Although further experiments need to be conducted, this situation could be explained by the fact that the tree canopy could work as a natural source of inoculum for A. niger. It is important to consider that A. niger does not produce aflatoxins, but some of its isolates can produce ochratoxins, another important concern for the nut industries in California.

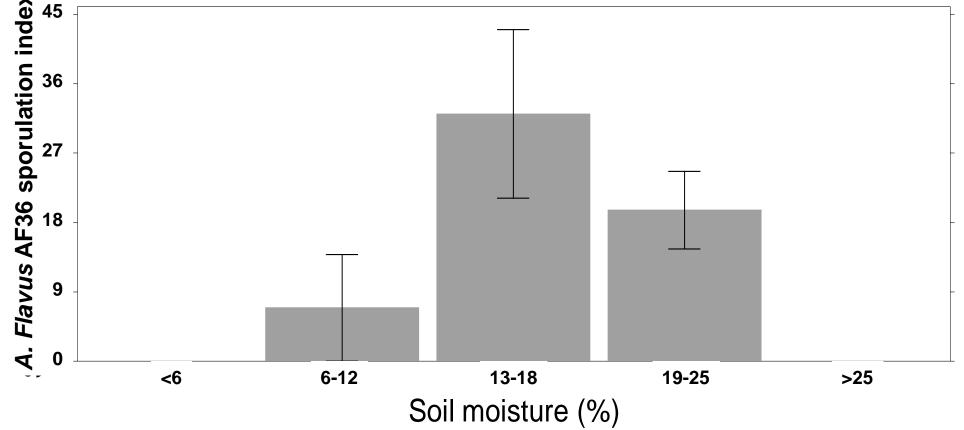
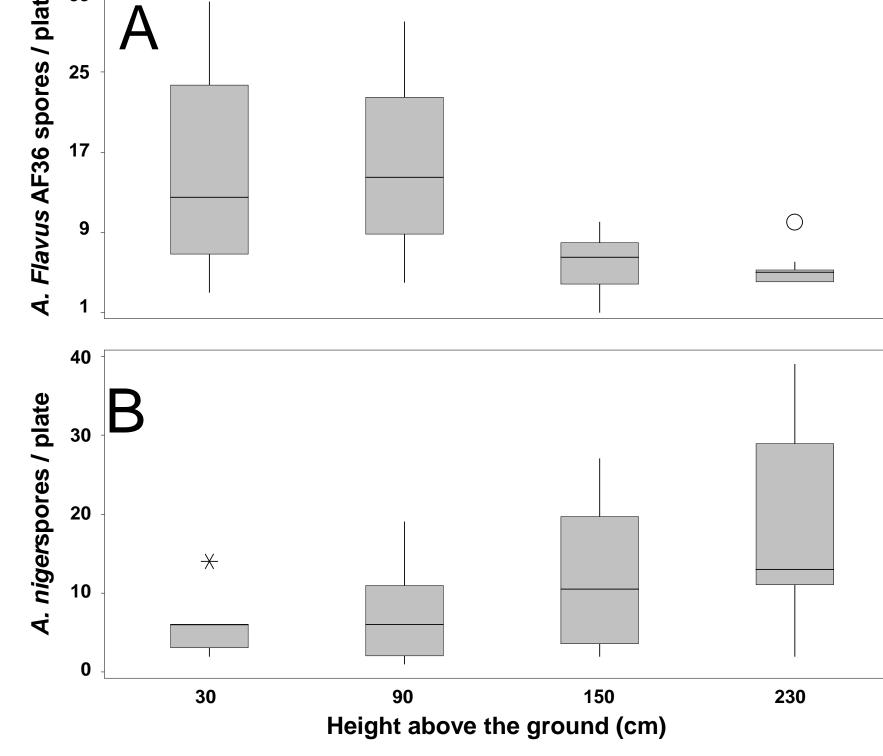


Figure 5. Effect of the soil moisture (%) on sporulation of *Aspergillus flavus* (AF36 Prevail<sup>®</sup>) biocontrol product grains.

Effect of soil water content on A. flavus AF36 sporulation. To determine the placement of the atoxigenic A. flavus biocontrol product grains for optimal sporulation, A. flavus AF36

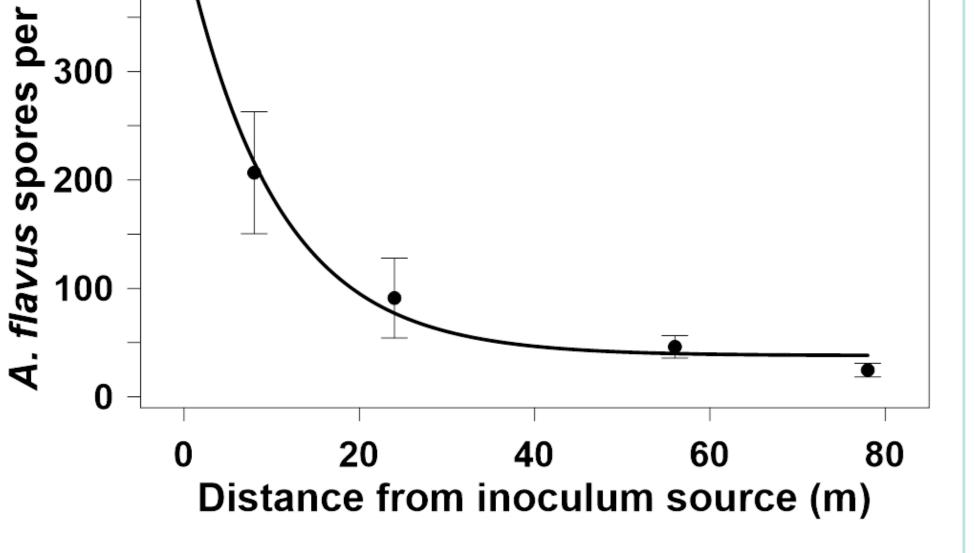


**Figure 4**. Box plots of **(A)** Aspergillus flavus AF36 and **(B)** A. niger spores per Petri dish at different heights above the ground (cm) where the biocontrol AF36 Prevail<sup>®</sup> was applied. The line within the box is the median. The top and bottom lines of the box represent 25 and 75<sup>th</sup> percentile of the data. Lines extending

Prevail<sup>®</sup> product grains were placed at different distances (from 25 to 250 cm) from the irrigation micro-sprinklers under field conditions. Both the density of sporulation of the A. *flavus* AF36 product grains and soil humidity by gravimetric analysis were periodically evaluated. In this experiment, we observed that there was an optimal sporulation of the AF36 product grains in the areas where soil moisture was between 13 and 18%. Conversely, AF36 sporulation was practically nonexistent in soil areas where there was excess (> 24% water content) or limited amount (<6% water content) of irrigation water (Fig. 5).

**Spore dispersal.** To determine the distance that the atoxigenic biocontrol fungus A. flavus AF36 is able to disperse from the source of inoculum in nut orchards, we applied the AF36 product, at a rate 10 times the normal, on the soil around one tree in the center of an orchard at Kearney Agricultural Research and Extension Center. Preliminary results indicate that the fungus is able to move at all directions. However, the population of total Aspergillus *flavus* decreased exponentially as distance from the spore source increased (Fig. 6).

Impact of arthropods on AF36 inoculum. The impact of different arthropods as potential cause of atoxigenic biocontrol product loss was evaluated under field conditions. A video camera (BirdCam, Wingscapes) was placed to monitor feeding behavior of arthropods on soils where the biocontrol product was applied. Results indicate that in no tilled soils, Oniscidea species (roly poly or pill bugs) and different ant species (Fig. 7) can impact the



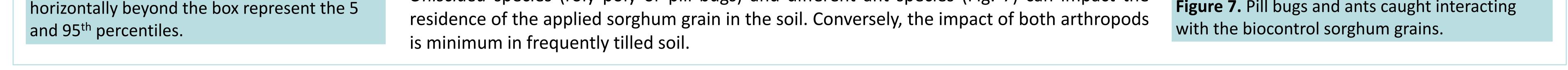
 $Y = 38.1 + 379.9 * e^{(-0.095X)}$ 

r<sup>2</sup> = 0.995

**Figure 6.** Number of spores of *Aspergillus flavus* per gram of leaf as a function of distance from the inoculum source.



Figure 7. Pill bugs and ants caught interacting





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