Project Number 76-G3
Project Title: Sex Pheromone Testing in Prevention of the Navel Orangeworm

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SUMMARY

The female sex pheromone of the navel orangeworm has been isolated and identified. Two forms (isomers) of the compound have been synthesized. Fóllowing confirmation of biological activity of the synthetic material work will begin on the development of a suitable formulation of the pheromone for orchard use.

Females call and release pheromone only during latter 1/3 of scotophase; pheromone not detectable(GC) on surface of pheromone gland during photophase and early scotophase.

B. PHYSIOLOGICAL VARIABLES

METHOD OF ANALYSIS

a.male age

bioassay

maximum male response from 2-4 day-old males; ca. 10 or more fold decrease in response among males older than 4 days; one-day-old males responded less well than did 2-4 day-old individuals but the difference appeared to be slightly less than 10 fold(i.e., on the border-line of the sensitivity of the bioassay).

b. female age

Gas Chromatography

no significant differences in quantity of pheromone from 1-4 day-old females; females older than 4 days generally did not call and were not analyzed re. pheromone content.

c. mating

bioassay

no sqnificant differences in pheromone response of males mated 24 h before bioassay and unmated males that could not be explained by age differences(see a above).

FEMALE Gas Chromat.

Erratic results; some mated females seemed to produce and release as much pheromone as did unmated individuals, but other females appeared to contain only about ½ as much pheromone after mating as did comparably aged virgin females.

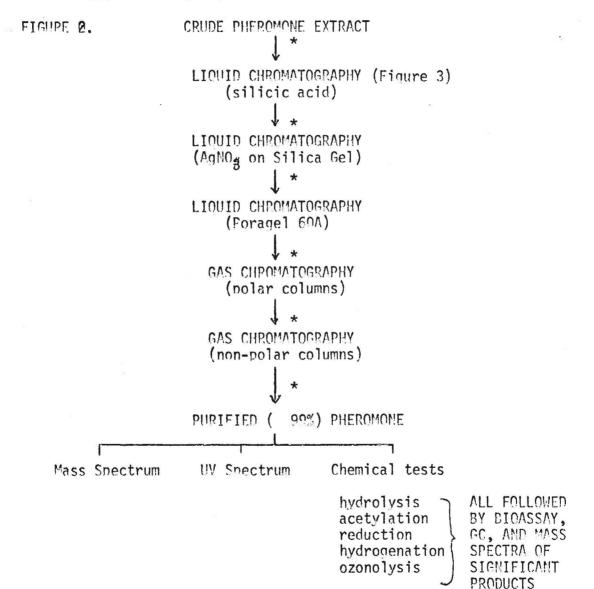
III. ISOLATION OF THE PHEROMONE

A. Influence of female or extract source on quantity of pheromone obtainable from NOW females

Table 1 summarizes the results of attempts to quantitatee obtainable pheromone from field and lab. females as well as pheromone from different sources of lab females.

SOURCE OF FEMALES	SOURCE OF EXTRACT	na PHEROMONE/ FEMALE	METHOD OF AMALYSIS
laboratory	filter paper	less than 0.5	bioassay
laboratorv	abdominal tip	ca. 1.0	GC & bioassay
laboratory	gland dips	0.8-1.7	GC & binassay
field	aland dins	2-4	GC & bioassay

B. Farly work had shown the the quantity of pheromone obtainable by the gland dip technique was at least as high as that obtained by more rigorous methods. Pip extracts had the additional advantage of being "clean" in the pheromone area on both polar and non-polar gas chymatographic columns. Despite the limitation of obtaining pheromone for only about 3 hours/ day, dip preparations were used as a pheromone collection method starting point for the following isolation procedure:



I. BIOASSAY--Pheromone extracts were prepared from calling NOW females using the method of Sower et al. (1973). Serial dilutions of the crude extract were bioassayed using the method and apparatus of Sower et al.(1973b), in which both male activation and orientation (attraction) were used as response criteria. In addition, the extracts were bioassayed using isolated males in a still air olfactometer (5 dram vial). Activation(and wing buzzing) were considered evidence of a positive response in this test.

The results of this series of bioassays are summarized in Fig. 1. The quantitative nature of the bioassay is evident, and the sensttivity of the assay is similar (in terms of female equivalents (FE)) to that which has been reported for a number of moth species. The orientation response in the wind tunnel and the activation response in still air were subsequently used as response criteria while monitoring isolation of the pheromone. Details of the bioassay procedure will be supplied upon request.

II. BIOLOGICAL STUDIES--Following is a brief summary of experiments that were conducted to determine the influence of environmental and physiological variables upon male response to and female production of the pheromone. As above, detailed methods will be supplied upon request.

A. Environmental variables

method of analysis

a. light

bioassav

Maximum male responsiveness at approximately full moonlight; there was a 10-50 fold decrease in male responsiveness with a 10 fold increase in light intensity.

b. temperature

bioassay

Maximum male responsiveness at 68-76°F; male response decreased at temperatures above 78°F. Lower temperatures (less than 68°F, were not tested).

c. time of day

bioassay

MALE RESPONSE

maximum male response during latter half of 10 h scotophase; greater than 1000 fold decrease in male responsiveness during photophase

MATTNG

OBSERVATION

that was extractable from the adult and which elicited sexual stimulation in the male. The limited quantities of pheromone that were available for isolation purposes materially slowed the identification of the compound. Isomers of the synthetic pheromone will be available for laboratory testing in January 1977. Extensive laboratory testing of the pheromone will begin by February 1977. Hopefully we will be able to come up with a suitable formulation of the material that will allow for field tests of the material during the 1977 crop year.

Peferences cited

- Sower, L.L., J. A. Coffelt and K. W. Vick. 1973. Sex pheromone: A simple method of obtaining relatively pure material from females of five species of moths. J. Econ. Entomol. 66: 1220-1222.
- Sower, L. L., K. W. Vick and J. S. Long. 1973. Isolation and preliminary biological studies of the female-produced sex pheromone of <u>Sitotroga cerealla</u> (Lepidoptera:Gelechiidae). Ann. Entomol. <u>Soc. Amer.</u> 66: 184-187.

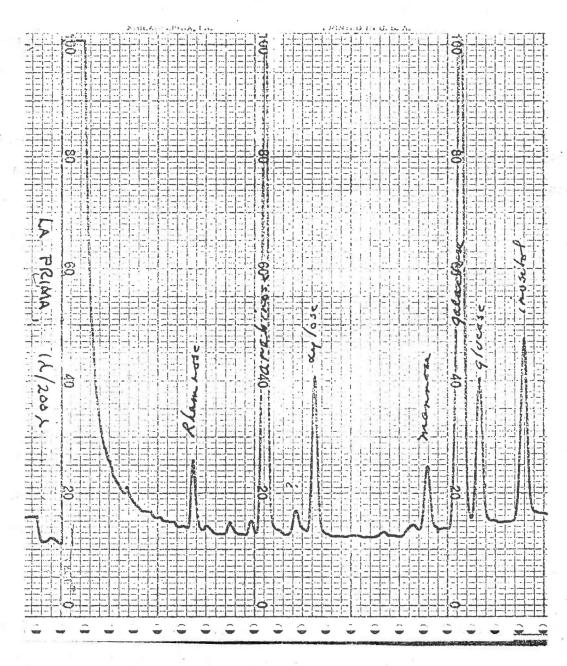


Fig. 3. Gas chromatographic tracing of sugar derivatives obtained from La Prima almond gum after hydrolysis with trifluoroacetic acid and subsequent reduction and acetylation with acetic anhydride.