Data-driven physiological modeling of canopy photosynthesis for precision irrigation management

Project Cooperators and Personnel:

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Grantee(s) of the Almond Board are REQUIRED to address sections A through G. These should be **submitted in PDF**, using Arial font size 12 for the main text, and be five to seven pages in length.

A. Summary (In laymen's terms – emphasize key findings and recommendations)

We sought to determine how canopy photosynthesis is affected by irrigation, to test the hypothesis that irrigation scheduling could be improved to increase total carbon gain for a given total irrigation input. The rationale is that growth and yield require carbohydrates from photosynthesis, which can be decoupled from water loss by variations in VPD, temperature and water deficit effects on the biochemistry of photosynthesis. Photosynthesis also typically sees "diminishing returns" from increasing transpiration, such that under heavy irrigation, it may be possible to reduce water applications with no detrimental impact on photosynthesis.

To test these ideas, we used sap flow and meteorological measurements to determine canopy conductance, and combined that data with measurements of the response of leaf photosynthesis to CO₂ and light to simulate canopy photosynthesis over a summer growing season, in two varieties (Aldrich and Nonpareil) in a research orchard in the northern Sacramento Valley near Arbuckle. Half of the study trees were stressed by withholding water for 7-10 days at two times during the season. Control trees were moderately stressed prior to harvest by reducing irrigation by 20%. Otherwise, all trees were irrigated to ETc.

We found that (1) photosynthetic biochemistry was unaffected by soil drought sufficient to cause nearly total closure of leaf stomata; (2) variations in VPD, but not temperature, caused a mild degree of decoupling of photosynthesis from transpiration; (3) in this orchard, photosynthesis was nowhere near saturated with respect to water loss, such that (3a) yield declined in the stress treatment approximately in proportion to the estimated reduction in canopy photosynthesis caused by stomatal closure during stress conditions, and (3b) optimal redistribution of a given amount of irrigation over the season would have little impact on total carbon gain; and (4) even mild water stress such as associated with 20% reduction in irrigation prior to harvest may cause substantial decline in water transport (hydraulic conductance), which has the potential to suppress photosynthesis even after resumption of full irrigation.

We conclude that irrigating to ETc during the main growing season is fairly efficient in terms of carbon gain, but that withholding water around harvest may substantially harm tree health and reduce current-year and future yield. The latter issue requires more intensive study.

B. Objectives (300 words max.)

- 1. Specify the goal(s) and specific objectives of the proposal if a collaborative effort, identify who is the lead for each objective
- 2. Identify annual outputs or milestones for each of the objectives

Main Goal: Develop and deliver a method for irrigation scheduling based on physiological modeling of canopy photosynthesis

B. Annual Results and Discussion (This is the core function of this report)

- 1. Describe activities and outputs for each objective
- 2. Discuss significance of these in terms of progress toward goals, change in approach, next steps or other conclusions based on this year's results

This was originally designed to be a 2-year project (2019-2021), and the objectives and milestones described in Section B were organized accordingly. It was subsequently announced that all current projects would terminate in 2020. We therefore sought to gather as much knowledge as we could within the available timeline.

Objective (1): Parameterize a canopy photosynthesis model in Nonpareil and Aldrich and drive it with continuous measurements of canopy conductance from sap flow.

We completed this objective and the first two milestones in November 2019. We will be unable to complete a 2nd year of parameter estimation (the third milestone originally listed for this objective) due to early termination of the project.

We successfully parameterized a canopy photosynthesis model for both Nonpareil and Aldrich (Figure 1) and drove the model using continuous measurements of canopy conductance from sap flow. We found no differences in photosynthetic parameters, nor their response to imposed water stress, in the two varieties, so we combined data across the two varieties.

Objective (2): To help scale the model from leaf to orchard level, quantify variation in PAR within tree crowns, and in intrinsic water use efficiency (iWUE, the ratio of photosynthesis to stomatal conductance) among trees.

We will collect sapwood cores in mid-winter to quantify variation in intrinsic water use efficiency among trees at the Nickels orchard using isotopic techniques. The samples will be processed in spring 2020.

We were unable to secure the instruments needed to measure canopy light penetration directly during summer 2019. These were to be borrowed from PI Buckley's former lab in Australia, but they were loaned to another party. We recently located another source for these sensors and will install them in early summer 2020.

Figure 2. Trends in summer gas exchange, showing declines in canopy and hydraulic conductances and photosynthes is in response temporary partial or total drought.

Objective (3): Determine how changes in irrigation affect modeled canopy photosynthesis and measured SWP as environmental conditions vary across a growing season.

We completed this objective in November 2019. We will be unable to complete a 2nd year of measurements (the second milestone originally listed) due to early termination of the project.

We found evidence of strong and rapid coupling between irrigation and both canopy conductance and simulated canopy photosynthesis (Figure 2). This held true for both pulses of regular irrigation (every 3 or 4 days at Nickels) and sustained reduction or withholding of irrigation. We also found evidence of strong decline in hydraulic conductance (the rate of water use that can be sustained for a given water potential gradient) during even moderate drought (Fig 2, lower panel). During sustained and repeated drought, these declines appeared to be irreversible. In both cases, our evidence suggests, although inconclusively, that hydraulic decline exacerbated the negative influence of drought on both stomatal opening and photosynthesis. Effects were substantial even in the control treatment, which experienced only a 20% reduction in irrigation during the harvest period.

Objective (4): To allow canopy photosynthesis to be modeled without sap flow data, parameterize and test a physiological model for canopy conductance in Nonpareil and Aldrich.

We found that the physiological model was not capable of adequately reproducing dynamics of sap flow with soil water potential interpolated linearly between measurements of pre-dawn stem water potential taken every 4-5 days (Figure 3). This was likely due to the frequent fluctuations in soil moisture caused by periodic (every 3-4 days) pulses of irrigation, with significant drying occurring rapidly after each pulse due to the site's poor sandy/gravel soils. Changes in soil moisture could be incorporated in the model if they could be converted to soil water potential; however, when we attempted to simulate soil water potential from soil water content, we found that the inferred pre-dawn water potentials were invariably far higher than measured (e.g.,> -0.002 MPa simulated, vs. -0.5 to -1.2 MPa measured). This likely indicates that soil drying affected the soil-to-root hydraulic interface rather than the water potential of the bulk soil per se until soil water content was close to the permanent wilting point. To account for

these effects would require much more intensive characterization of the plant hydraulic vulnerability curve (the response of hydraulic conductance to water potential, and particularly the soil-root interface component), which was beyond the scope of this study.

Objective (5): To determine guidelines for more effective irrigation scheduling, use the model to simulate the impact of a range of potential irrigation strategies.

Although this objective was planned to be carried out during spring-summer of 2020, we completed it for the 2019 season's data during fall of 2019. Because this objective also depended upon the physiological conductance model from Objective (4), which we found to be inadequate, we utilized a different approach. We characterized the overall

Figure 3. Performance of physiological model for canopy conductance with daily pre-dawn water potential interpolated from measurements every 4-5 days. Lines = best fit (control: $y = 0.46x + 3.18$, r^2 = 0.46; stress: y = 0.53x + 2.02, r^2 = 0.52).

relationship between canopy photosynthesis and canopy transpiration (Figure 4), and then used that relationship to simulate the impact of redistributing water loss in time over the growing season in the manner that would maximize simulated wholeseason canopy photosynthesis. The basic rationale is that it is less efficient to irrigate under some conditions than others, because differences in VPD, temperature and other factors can decouple water loss from photosynthesis.

Somewhat to our surprise, we found that redistribution of water use could, at best, improve whole-season carbon gain by about 5%. Even that was only possible if we assumed that an irrigation strategy to achieve this could be designed and implemented with perfect precision, and that the

resulting shifts in plant water status would have no other detrimental impacts. In reality, our field data suggest that even minor water stress during the season, such as in response to withholding of water around harvest, can result in substantial and possibly irreversible reduction of plant hydraulic conductance (Fig 2).

Consistent with those results, we also found that yield declined substantially in the stress treatment, and approximately in proportion to the estimated reduction in total photosynthesis caused by the stress treatment (Figure 5, left). We also estimated the reduction in photosynthesis caused by mild harvest stress in the control treatment to be approximately 23%

Figure 5. Yield and photosynthesis for control and stressed treatments (left; grey symbol at right), and the estimated decline in photoynthesis caused by reduction of irrigation prior to harvest in the control treatment (red line at right).

There are two important lessons from this result. One is that managing canopy transpiration by irrigating to ETc is actually quite efficient with respect to canopy photosynthesis. The factors that we expected to drive substantial decoupling of photosynthesis from transpiration were not as important as we had anticipated. The other lesson is that even mild stress such as that experienced during a typical harvest period (in this case, the farm managers imposed a 20% decline in irrigation inputs beginning three weeks before harvest) may lead to damaging loss of water transport capacity, which would impact late-season photosynthesis and growth and possibly performance in subsequent years.

Objective (6): Develop a stand-alone package, including a user-friendly app, to provide irrigation advice to growers based on modelling canopy photosynthesis.

This work was intended to be carried out during the final year of the project, and subject to useful insights emerging from the first year's analysis. Since the project will be terminated at the end of the current project year, and furthermore since our analysis found that irrigating to ET is actually fairly efficient with respect to photosynthesis, we have not pursued Objective 6.

C. Outreach Activities

1. Please describe outreach activities including the event description (date, location, topic of the presentation, approx number of participants and type of audience)

Results of this project to date were presented at the Almond Conference in Sacramento on 12 December 2019, to approximately 100 participants of varying backgrounds. We also presented a poster summarizing the results at the Almond Conference.

D. Materials and Methods (500 word max.):

1. Outline materials used and methods to conduct experiment(s)

2. Note any challenges or unforeseen developments that were encountered resulting in change of methodology, timeline, or scope of project

Setting and treatments. We studied 20 individual trees at the Nickels Soil Laboratory near Arbuckle, CA, of which 10 were Nonpareil and 10 Aldrich. Trees were irrigated to ETc once every 3-4 days unless otherwise noted. Irrigation was withheld from 5 trees of each variety ("stress" treatment) at each of three times during the season, for 7-10 days (early June, early July and early August). The remaining 5 trees of each variety (the "control" treatment) were irrigated normally until three weeks prior to harvest, when irrigation was reduced to 20% below ETc, and then withheld entirely from 4 days before to 6 days after shaking.

Sap flow. We measured sap flux (sapwood water flow per unit of cross-sectional sapwood area) in each tree using the double-ratio method (DRM). Four needles (one heated needle and three temperature sensors, located 7.5 mm below, 7.5 mm above and 22.5 mm above the heater, respectively; each needle was 1.27 mm in diameter and 30 mm in length) were installed in each tree trunk approximately 60 cm above the ground and underneath the bark, and then insulated by wrapping the tree with batting insulation and mylar-coated bubble wrap (Reflectix). The DRM method was described in previous project reports for the ABC, and has been validated by lysimetry and tube flow experiments.

Canopy conductance. We calculated canopy conductance by dividing sap flow by estimated leaf to air water vapor mole fraction gradient (computed from air VPD) and then empirically adjusting the result to match mid-day values of stomatal conductance measured directly on each tree in early July.

Canopy photosynthesis. We measured photosynthetic parameters in each tree before and after the July dry-down, by recording the response of leaf net $CO₂$ assimilation rate to intercellular CO₂ concentration and fitting the biochemical photosynthesis model of Farquhar et al. (1980) to the results. We used previously published temperature responses for these parameters in almond. We combined these parameters with measurements of light intensity and temperature from an adjacent tower and canopy conductance to calculate leaf photosynthesis of sunlit leaves, and then scaled those values to the canopy used the sun/shade scaling method of de Pury and Farquhar (1997), based on leaf area index inferred from canopy light penetration measurements from Bruce Lampinen's mobile light bar.

Water re-distribution modeling. We generated a polynomial relationship between daily canopy photosynthesis and transpiration for July-August 2019 from the measurements and modeling described above, and then used this relationship to predict seasonal photosynthesis with daily transpiration adjusted while maintaining seasonal transpiration constant. The adjustments consisted of reducing water loss on days of high water loss (which are associated with lower marginal efficiency with respect to photosynthesis) and increasing it on days of low water loss.

E. Publications that emerged from this work

- 1. List peer review publications in preparation, accepted or published
- 2. Other publications (e.g. outreach materials)
- 3. Please provide copies of publications

Nothing to report.